

High-resolution Chromosome Conformation Capture: Hi-C

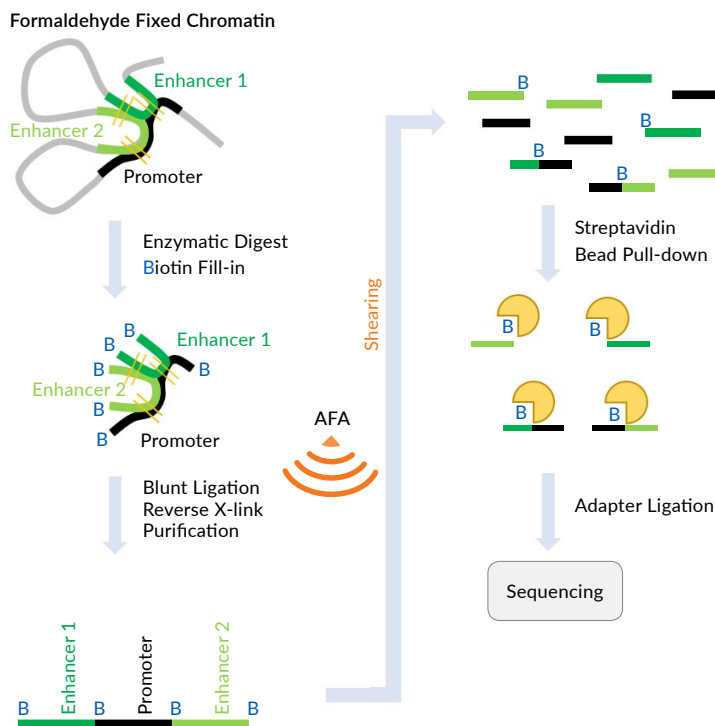
Scientific Relevance

- Three-dimensional chromatin organization regulates gene expression ¹
- Aberrant chromatin looping causes altered gene regulation in malignancies including solid tumors as well as hematologic neoplasms ²
- Characterization of 3D-chromosomal conformations allows classification of cancer subtypes ³
- Cancer progression can be alleviated by inhibiting certain chromatin loop formations ^{4,5,6}
- Hi-C provides a powerful tool to better characterize 3D chromatin organization and helps to uncover the impact of cancer risk-associated SNPs ^{7,8}

Challenges

- Protocol requires several replicates to retrieve reliable 3D-conformation data, good sets of controls, and optimizations are essential
- Unbiased, reproducible shearing with a tight DNA fragment size distribution is required to capture all chromosomal interactions especially in low input derivatives ² of the method

Workflow



Schematic representation of Hi-C workflow adapted from [Davies et al.](#)

Crosslinked chromatin is digested and the overhangs of the restriction enzymes are filled-in using biotin-labelled nucleotides following blunt-end ligation. Reproducible shearing followed by Streptavidin-bead pull-down allows the efficient and selective purification of chimeric DNA ligation junctions which are subjected to sequencing.

Advantages of Adaptive Focused Acoustics® (AFA®)

[AFA technology](#) is a very gentle, reproducible, and tuneable shearing method.

- Random shearing guarantees an unbiased fragmentation of ligation products
- The tight size distribution ensures comprehensive representation of all ligation junctions in the sequencing library
- Reproducible shearing allows reliable comparison of samples from different origins such as cancer subtypes or different stages of progressive diseases

Suggested Covaris Products

- [Covaris Focused-ultrasonicator](#) (M-Series, S-Series, E-Series, or LE-Series)

Citations

- [Van Berkum et al. Hi-C: a method to study the three-dimensional architecture of genomes. *Vis Exp.*, \(2010\)](#)
- [Ramani et al. Mapping three-dimensional genome architecture through in situ DNase Hi-C. *Nat Protoc.*, \(2016\)](#)
- [Elphege et al. Targeted degradation of CTCF decouples local insulation of chromosome domains from genomic compartmentalization. *Cell*, \(2017\)](#)